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Anticancer and antimicrobial properties of extracted bioactives of Rosemary (*Rosmarinus officinalis* L. syn., *Salvia rosmarinus* Spenn.)

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Abstract

Rosemary is one of the potential herbs being exploited by various industries worldwide. It has been used in the treatment of cancer, in aromatherapy, and psychological treatments, and as an antimicrobial agent. The major constituents of the essential oil are 1,8-cineole, a pinene, camphene, β pinene, carnosol, and carnosic acid which are attributable to all the important properties of the essential oil. Many novel extraction procedures such as microwave assisted steam distillation, multistage steam distillation, CO₂ supercritical extraction, microwave hydrodiffusion and gravity, solvent free microwave extraction, controlled instantaneous decompression have emerged apart from the conventional hydrodistillation and steam distillation, by realizing the advantages over the traditional ones. Hence, this present study focuses on novel and profitable extraction techniques, chemical constituents, anticancer and antimicrobial properties of *Rosmarinus officinalis* L.

1. Introduction

Medicinal and aromatic plants have recently gained a wide scope to be exploited for their valuable essential oils, extracts for varied purposes like additives in food, plant protection, and also for their research interest due to their bioactive secondary metabolites, which offer promising medicinal applications (Mathe, 2018; Sokhibjon *et al.*, 2024).

Rosemary (*Rosmarinus officinalis* L. syn., *Salvia rosmarinus* Spenn.) is one of the important herbs belonging to the family Lamiaceae. It is an evergreen perennial shrub grown across the world for its health benefits and therapeutic potential derived from its oil. It is also referred to as the “Dew of the sea” due to its aroma and fragrance (Selavaraj *et al.*, 2022). The leading exporters of rosemary are India, Turkey, and South Africa. India exports rosemary to the United States, Australia and Germany. From India, the yearly export is gaining an increase of 3% (Volza Grow Global, 2024). Besides the whole plant, there is a growing demand for the oil and extract of the herb. This has been used anciently by the tribes in treating splitting headaches, relieving muscle spasms, and in psychotherapy (Abu-Al-Basal, 2010; Rahbardar and Hosseinzadeh, 2020; Manville *et al.*, 2023). This herb is noticed for its unique taste and pleasant smell. It not only possesses antibacterial and anti-fungal properties but also

has beneficial effects on human health. Its potential is being exploited across various industries and pharmaceuticals (Pawowska *et al.*, 2020). Cosmetics is one of the growing industries, that explores this valuable plant to treat cellulite, as an antioxidant to reduce ageing and can be used in the treatment of skin cancers (Macedo *et al.*, 2020). As it contains polyphenols such as carnosic and rosmarinic acid, its use has also been exploited in treating type 2 diabetes in humans (Naimi *et al.*, 2017). Rosemary extracts, essential oil, and hydrosols have been found to have antimicrobial (Abozid and Asker, 2013), antinociceptive (Beltrán-Villalobos *et al.*, 2017), anticancer (Jaglanian *et al.*, 2020a), antioxidant (Saini *et al.*, 2020), antidiabetic (Kabubii *et al.*, 2024), antidepressant (Guo *et al.*, 2018), anti-inflammatory (Habtemariam, 2023), antispasmodic (Ai-Sereiti *et al.*, 1999) properties. It is also used to enhance memory and increase sleep quality (Nematollahi *et al.*, 2018). The evaluation of essential oils after the process of distillation, are separated and analyzed majorly through gas chromatography (GC) or gas chromatography mass spectrometry (GC-MS) (Pintore *et al.*, 2002; Tschiggerl and Bucar, 2010). Thorough GC-MS, 15 to 80 chemical constituents have been identified after distillation procedures (Socaci *et al.*, 2009; Rajeswara Rao *et al.*, 2010; Amin *et al.*, 2017; Abozid and Asker., 2013). The major constituents identified were α -pinene, 1,8-cineole, camphor, verbenone, borneol, linalool, eucalyptol, and borneol under various conditions and growth factors (Gachkar *et al.*, 2007; Tschiggerl and Bucar, 2010; Hcini *et al.*, 2013; Tomi *et al.*, 2016; Mwithiga *et al.*, 2022). The essential oil consists of neutral lipids, monounsaturated, polyunsaturated, and saturated fatty acids, oxygenated monoterpenes, monoterpene hydrocarbons, tocopherols, and phenols (Tigrine-Kordjani *et al.*, 2012; Elbanna *et al.*, 2018; Doozakhdarreh *et al.*, 2022;). The percentage of major constituents present in majority

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of the genotypes include 1,8-cineole (20.1 to 37.5%), camphor (12.35 to 13.55%), α pinene (8.58 to 11.8%), β pinene (6.7 to 6.10%), borneol (4.0 to 5.61%), camphene (5.07 to 5.85%), α terpineol (4.30 to 8.17%), limonene (3.19 to 3.04%), *p* cymene (2.42 to 3.11%), verbenone (6.20 to 8.14%) (Rajeswara Rao *et al.*, 2010; Tavassoli *et al.*, 2011; Hcini *et al.*, 2013; Amin *et al.*, 2017). Upon polyphenol analysis diterpenoids and flavonoids were the major classes. Carnosic acid was the predominant polyphenol present in it (Mena *et al.*, 2016).

Rosemary extracts have also been used in the food industry to preserve various natural food products through their antibacterial, antifungal, and antioxidant properties. Its use has been mainly exploited in the preservation of meat and meat products due to its aroma (Al-Hijazeen and Al-Rawashdeh, 2019; Lorenzo *et al.*, 2021; Boukhari and Fatimi, 2023).

Though plants in this world are part of curing diseases and in rural healthcare (Majeed, 2017) and in the modern world, the usage of chemical drugs and medication plays a major part in the treatment of all diseases and disorders. However, the awareness of the side effects and the chemical toxicity in humans has shown light upon traditional and herbal medicines. There is also a scope widened for combination treatments with medicinal plants (Rathore *et al.*, 2017). This review highlights the major constituents of rosemary extract, and the procedures to extract them. The anticancer effects on breast cancer, prostate cancer, colon cancer, and skin cancer have been presented. Antimicrobial effects for bacteria and fungi have also been discussed in this review.

2. Extraction procedures

Rosemary oil extraction carried out by steam distillation (SD) and hydrodistillation (HD) revealed that steam distillation is found to have more oil recovery than hydrodistillation. 1.2% of essential oil yield was obtained through steam distillation and 0.44% from hydrodistillation which was very low. 80% of the oil was recovered in 10 min in steam distillation rather than 30 min in hydrodistillation

(Boutekdjiret *et al.*, 2003; Presti *et al.*, 2005; Arafa, 2019). The maximum yield of oil was obtained through steam distillation. Spadi *et al.* (2021) stated that the optimum time for greater oil yield was 120 min, with no increase in yield after this point.

Multistage steam distillation proved advantageous for industrial applications. Steam had more contact time with the leaves, so the yield obtained was also greater in a shorter duration of 30 min, 85% of the oil was recovered through multistage steam distillation. (Malekydozadeh *et al.*, 2012).

Microwave assisted hydrodistillation (MAHD) is one of the new emerging methods in extraction procedures. Unlike hydrodistillation, this process is a solvent-free extraction procedure. The time taken for extraction is 20 min. The oil yield and the characteristics are like those obtained from steam or hydrodistillation but had major advantages proving to be environmentally friendly by emitting less CO₂ into the atmosphere, decreasing energy consumption, and reducing the cost of production (Karakaya *et al.*, 2014; Elyemni *et al.*, 2019; Ferreira *et al.*, 2020). By increasing the microwave generation by 50%, more oxygenated compounds are obtained with decreased monoterpenes. Hence, the obtained oil quality was superior under microwave assisted hydrodistillation (Fazlali *et al.*, 2015).

CO₂ supercritical extraction gives greater yields of essential oil than hydrodistillation and steam distillation. The essential oil extracted through this method shows 14 times greater antioxidant activity than that obtained through other methods of extraction (Conde-Hernández *et al.*, 2017). Superheated water which is about 125% and 175% under pressure was found to extract oxygenated aroma compounds, but this leaves behind some monoterpenes, higher hydrocarbons, lipids and waxes. The oxygenated compounds obtained are more when compared to steam distillation (Basile *et al.*, 1998). Microwave hydrodiffusion and gravity (MHG), proves to have many advantages of using no water or solvent which is environmentally safer, reducing the operational time from 3 h of hydrodistillation to 15 min, with lesser energy costs and essential oil with more antimicrobial and antioxidant activity (Bousbia *et al.*, 2009).

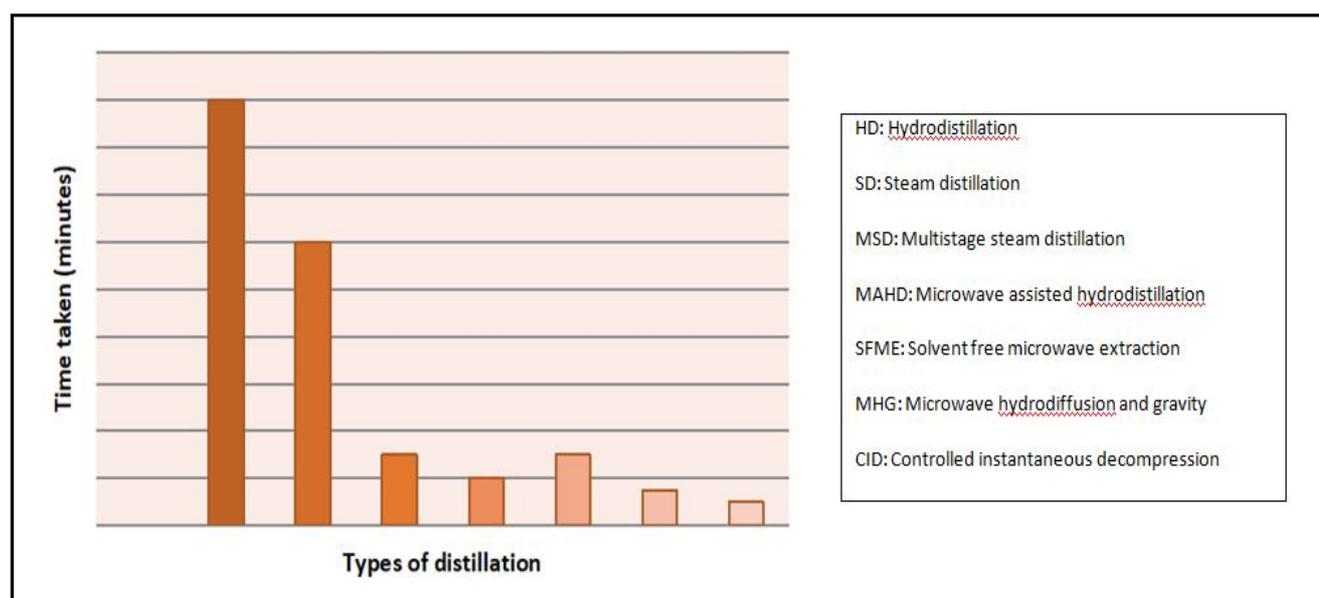


Figure 1: Distillation types and time taken.

Solvent free microwave extraction (SFME) is a combination of microwave heating and dry distillation. The yield and oil quality were the same in both the solvent free microwave extraction and hydrodistillation, except for the time taken (30 min and 180 min). SFME is found to be an alternative environmentally friendly technique to be used in industries for extraction (Filly *et al.*, 2014). Enzyme assisted extraction with cellulase and hemicellulase and in combinations gave a higher yield of essential oil (Hosni *et al.*, 2013). Controlled instantaneous decompression, an emerging method, can be used to extract 97% of essential oil in about 10 min by applying a steam pressure of about three bar (Rezzoug *et al.*, 1998). Subjecting rosemary leaves with a moisture content of 0.40 g H₂O/g of dry material, subjecting to a short period of steam pressure of 410 kPa and then dropping pressure instantaneously to vacuum to 50 kPa, with an operational time of 16 min is effective in extracting 96% of essential oil through polynomial equation (Rezzoug *et al.*, 2005).

3. Classification of essential oil

Rosemary oil is classified into four categories based on the major component. These variations are found in different genotypes, varied agroclimatic conditions, and differences in extraction methods. In India, the essential oil is cineoliferous and camphoriferous type (Rajeswara Rao *et al.*, 2010; Borges *et al.*, 2019; Sadeh *et al.*, 2019) (figure 2).

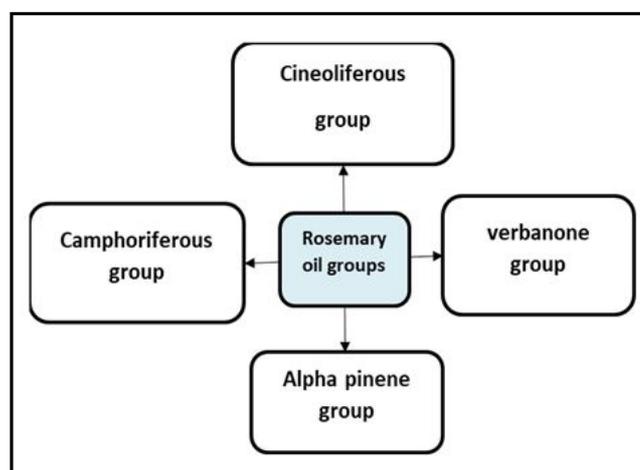


Figure 2: Rosemary essential oil based on the major component present.

The components of the extracts are classified into various classes. This may include groups like monoterpenes, monoterpene hydrocarbons, oxygenated monoterpenes, sesquiterpenes, diterpenes, phenolic acids, and flavonoids (Almela *et al.*, 2006; Borges *et al.*, 2019).

Table 1: Classes of predominant constituents present in rosemary

Class	Components	Chemical formula	Reference
Monoterpenes	α -pinene	C ₁₀ H ₁₆	Borges <i>et al.</i> , 2019
	Camphene	C ₁₀ H ₁₆	Kadri <i>et al.</i> , 2011
	Verbenone	C ₁₀ H ₁₄ O	Borges <i>et al.</i> , 2019
	α -thujone	C ₁₀ H ₁₆ O	Kadri <i>et al.</i> , 2011
Monoterpene hydrocarbons	Limonene	C ₁₀ H ₁₆	Borges <i>et al.</i> , 2019
	β -pinene	C ₁₀ H ₁₆	Yosr <i>et al.</i> , 2013
	terpinolene	C ₁₀ H ₁₆	Borges <i>et al.</i> , 2019
	β -myrcene	C ₁₀ H ₁₆	Yosr <i>et al.</i> , 2013
	<i>p</i> -cymene	C ₁₀ H ₁₄	Borges <i>et al.</i> , 2019
	Oxygenated monoterpenes	1,8-cineole	C ₁₀ H ₁₈ O
Camphor		C ₁₀ H ₁₆ O	Tavassoli <i>et al.</i> , 2011
Borneol		C ₁₀ H ₁₈ O	Mena <i>et al.</i> , 2016
Linalool		C ₁₀ H ₁₈ O	Borges <i>et al.</i> , 2019
Phenolic acids		Rosemarinic acid	C ₁₀ H ₁₆ O ₈
	Hydrobenzoic acid	C ₇ H ₆ O ₃	Mena <i>et al.</i> , 2016
Sesquiterpenes	Caryophyllene	C ₁₅ H ₂₄	Borges <i>et al.</i> , 2019
	α -humulene	C ₁₅ H ₂₄	Kadri <i>et al.</i> , 2011
Flavonoides	Apigenin	C ₁₅ H ₁₀ O ₅	Almela <i>et al.</i> , 2006
	Lutenin	C ₄₀ H ₅₆ O ₂	
Diterpenes	Carnosic acid	C ₂₀ H ₂₈ O ₄	Almela <i>et al.</i> , 2006

4. Anticancer properties

The essential oil of rosemary is found to have no cytotoxic effects on cells and hence can be used in the treatment of cancer (Al-Maharik *et al.*, 2022). The three main active ingredients found in rosemary extract carnosol, carnosic acid, and rosmarinic acid have been found to increase the effectiveness of CDDP (cis diamminedichloroplatinum), a drug used in cancer treatments (Tai *et al.*, 2012). Chloroformic rosemary extract and ethanolic oats extract bring about condensation of chromatin, which leads to cell death (Alzaharna *et al.*, 2020). Carnosic acid in rosemary extract showed the most effective control of cell multiplication (13 to 30%), with a 19 μM dosage concentration within 48 h of treatment (Yesil-Celiktas *et al.*, 2010).

4.1 Colon and prostate cancer

The components such as carnosic acid, carnosol, 1,2-methoxy carnosic acid, taxodione, and betulinic acid are responsible for the anti-proliferative effects in human colon cancer (Borrás-Linares *et al.*, 2015). Ethanolic rosemary extract had growth inhibition effects against the growth of Caco-2 lines of colon cancer cells (Alzaharna *et al.*, 2020). Administration of rosemary extracts *in vitro* for colorectal cancer increased the reactive oxygen species leading to death of cancer cells. Oral administration in athymic nude mice with grafted human colon cancer cells reduced tumor formations (Pérez-Sánchez *et al.*, 2019).

It has been found that rosemary essential oil prevents proliferation, survival, and the spread of the androgen sensitive PC 3 and 22VR1 prostate cancer cells. Significantly, there is no impact of rosemary essential oil on the normal epithelial prostate cells (Jaglanian *et al.*, 2020b). Reduced cell viability was observed by testing rosemary against prostate cancer cells LNCaP with IC_{50} values (Bourhia *et al.*, 2019). *In vitro*, administration of carnosol has been shown to inhibit the G2 cycle and cyclin production. Carnosol targets the AMPK pathway and brings down cancer cell proliferation (Johnson *et al.*, 2008). Testing five rosemary extracts with high carnosic acid in both *in vivo* and *in vitro* conditions has confirmed that the combination of carnosic acid with carnosol was more effective than their individual effects (Dilas *et al.*, 2012). Rosemary extract had a similar effect on HDAC 2 as SAHA suberoylanilide hydroxamic acid which is used in PCa cancer treatment, which brings about apoptosis of cell lines (Jang *et al.*, 2018)

4.2 Breast cancer

Extract with the highest amount of carnosic acid, 42.2% (Inolens50) is found to have a maximum antiproliferative effect against breast adenocarcinoma (González-Vallinas *et al.*, 2014). Rosemary extract showed moderate antiproliferative effects on cancer cells and with Bleomycin, but radio-mimetic bleomycin treatment showed better results on breast cancer cells (Mrđanoviæ *et al.*, 2019). Carnosol in rosemary extracts has been found to alter chromatin structure in MDA-MB-1231 and Hs578T breast cancer cells which brought down the functional ability (Alsamri *et al.*, 2021). Rosemary extract has been found to reduce cell multiplication, activation of Akt, and mTOR signaling of MDA-MB-231 breast cancer cells (Jaglanian and

Tsiani, 2020). Essential oil at 500 $\mu\text{g}/\text{ml}$ was effective (81 to 89%) on MCF7 breast cancer cells (Hussain *et al.*, 2010). Petroleum ether extract had anticancer activities on MCF7 breast cancer cells and HCT 11 human colon cancer cell lines at IC_{50} rates of 3.77 $\mu\text{g}/\text{ml}$ and 3.09 $\mu\text{g}/\text{ml}$, respectively (Ali, 2021). Mrđanoviæ *et al.* (2019) stated that rosemary extract with the drug BLM (bleomycin) was more effective than their individual treatment.

4.3 Skin cancer

Application of methanolic extract from rosemary leaves has been found to prevent the tumor formation induced by the application of Benzo (α) pyrene [B (α) P], 7,12-diethylene (α) anthracene (DMBA) treatment in mice. The covalent bond formations between [B (α) P] and epidermal DNA causing the tumor was disrupted when 1.2 or 3.6 mg of rosemary extract was topically applied before [B (α) P] treatment (Wang *et al.*, 1994). Cattaneo *et al.* (2015) observed that 65% of the hydroalcoholic extract of rosemary controls human melanoma, A375 group of cells by cytological effects.

4.4 Treatment for other cancers

Treating NSCLC H1299 (nonsmall cell lung cancer) cells at IC_{50} 19 $\mu\text{g}/\text{ml}$ of rosemary extract showed antiproliferative cell properties, induced cleavage of PARP markers of apoptosis, and prevented cancer spread (Neill *et al.*, 2022). MDA-MB-231, triple negative human lung cancer cells that had no recovery with hormonal treatments, showed antitumor activities as there was increased cleavage of PRAP (proline rich acidic protein), leading to apoptosis (Jaglanian and Tsiani, 2020). Wang *et al.* (2012) compared essential oil and its constituents α pinene, β pinene, and 1,8-cineole on human ovarian cancer lines SKOV3, HO-8910, and Bel-7402 and found that the essential oil of rosemary had more anticancer properties than individual constituents.

Treatment with rosemary essential oils revealed that there was maximum control towards the cell multiplication in human pancreatic cancer cell lines SW1990 and gastric epithelial cancer cell line NCI-N87 (Huang *et al.*, 2023). Jang *et al.* (2018) suggested that when then cancerous keratinocytes were cultured with rosemary extract at a concentration of 1.25% helps in the control of cell multiplication of keratinocytes HaCaT by regulating cell death. Carnosic acid proved to be a potential agent in enhancing the effects of sestrin2 and MRP2 for increasing the effects of Nrf2 genes in HepG2 liver cancer cells (Tong *et al.*, 2017). In human non-small cell lung cancer lines H441, H520, and H661 the IC_{50} for carnosic acid through MTT(3-(4,5-dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide assay) method was found to be 20 μM , 40 μM , 40 μM for the lines respectively. In the cell lines, the controlled death expressions were elevated, by changes in the genes such as Bax, Bak, caspase-3, and p53 mediating apoptosis (Duran *et al.*, 2024). In cell lines H1299 and H460, carnosic acid as a potential anticancer agent inhibited cell multiplication and induced death through the control of over-expressed AMPK (adenosine monophosphate activated protein kinase) activity. Sestrin 2 involved in the expression of cancer activity was also controlled by the recycling process by degradation of the organelles (Neill *et al.*, 2024).

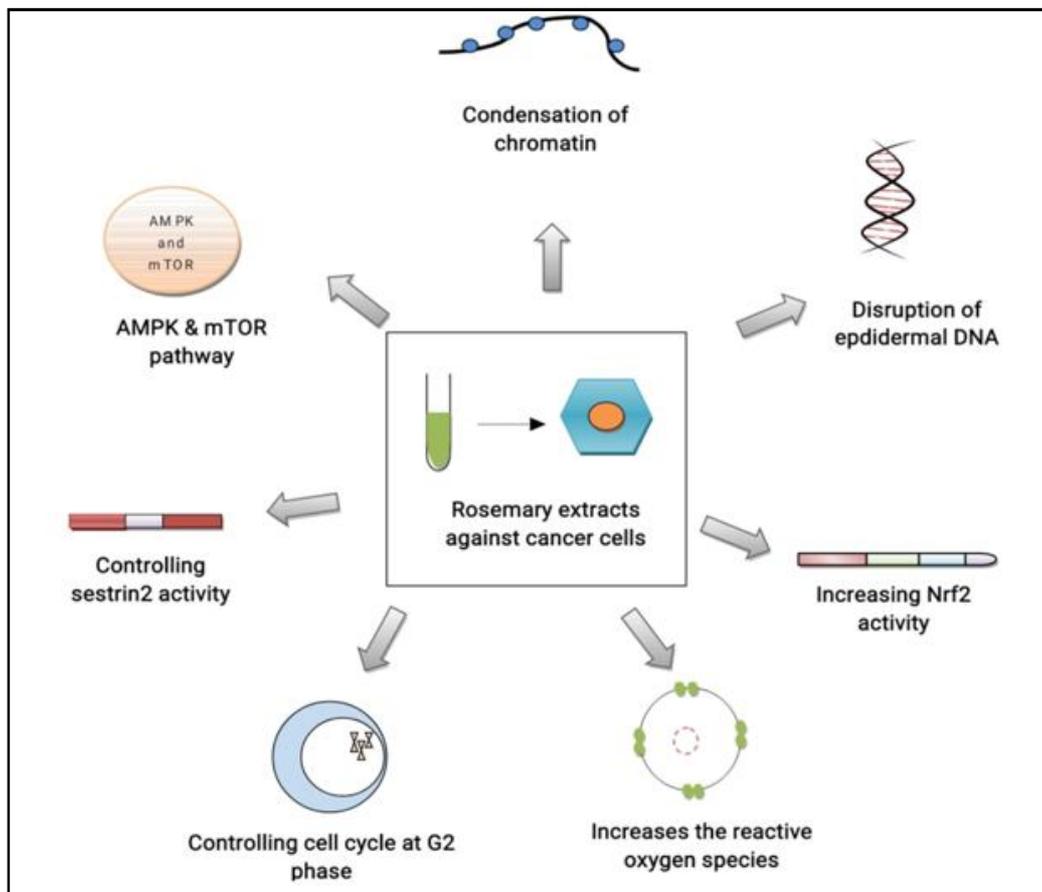


Figure 3: Mechanism of action of rosemary extracts against cancer cells.

Table 2: Anticancer properties of Rosemary extract and its constituents

Cancer type	Type of dose	Effect	Reference
Colon cancer			
Caco-2, colon cancer cell lines Colorectal cancer	Carnosic acid, carnosol, 1,2-Methoxy Carnosic acid	Antiproliferative	Borrás-Linares <i>et al.</i> , 2015
	Ethanollic extract	Growth inhibition	Alzaharna <i>et al.</i> , 2020
	Rosemary extract	Increases ROS and leads to death of cells	Pérez-Sánchez <i>et al.</i> , 2019
Prostrate cancer			
PC-3 and 22VR1 cell line	Rosemary essential oil	Prevented proliferation, survival and spread	Jaglanian <i>et al.</i> , 2020
LNCaP	Extract	Reduced cell viability	Bourhia <i>et al.</i> , 2019
	Carnosol administration	Inhibits G2 stage in cell cycle and reduces cell multiplication	Johnson <i>et al.</i> , 2008
HDCA2 cell line	Rosemary extract	Similar effect as Suberoylanilide hydroxamic acid	Jang <i>et al.</i> , 2018
	Carnosic acid+carnosol	Decreased cell proliferation	Dilas <i>et al.</i> , 2012

Breast cancer			
MCF-7	Rosemary extract + Bleomycin	Antiproliferative effects	Mrđanović <i>et al.</i> , 2019
	Carnosol	Alters chromatin structure	Alsamri <i>et al.</i> , 2021
	Rosemary extract	Activation of Akt and mTOR pathways	Jaglanian and Tsiani, 2020
	Petroleum ether extracts	Anticancer activities	Ali, 2021
Skin cancer			
A375 Non small cell lung cancer NSCLC Pancreatic SW1990 and NCI-N87 Gastric cancer Keratinocytes-HaCaT HepG2 liver cancer	Methanolic rosemary extracts	Prevents The covalent bond formation between [B (α) P] and epidermal DNA	Wang <i>et al.</i> , 1994
	65% hydro alcoholic extract	Cytological effects	Cattaneo <i>et al.</i> , 2015
	Rosemary extract	Showed anti proliferative cell properties, induced cleavage of PARP markers	Neill <i>et al.</i> , 2022
	Essential oil	Cancer control	Wang <i>et al.</i> , 2012
	Rosemary cell culture	regulation of cell death	Jang <i>et al.</i> , 2018
Carnosic acid	Increases the effects of Nrf2 genes	Tong <i>et al.</i> , 2017	

5. Antimicrobial activity

5.1 Antibacterial activity

Rosemary extracts have been found to have great antibacterial potential. The main components that cause the inhibitory action of bacteria are carnosol, rosmannol and isorosemanol. The inhibitory action of the extract is due to phenotypic and genotypic changes in the bacteria cells (Walid *et al.*, 2022). Rosemary oil as a whole proved to be a better antibacterial agent when compared to its major constituents like 1,8-cineole, α pinene, and β pinene when tested individually. Hence, the use of rosemary essential oil as a whole is widely preferred. Methanolic extracts also had a lesser effect than the actual extract (Abdoul-Latif *et al.*, 2021; Wang *et al.*, 2012).

The minimum value for bacterial growth inhibition (MIC) was 0.625 and 1.25% and the minimum antibacterial effect was 1.25 and 2.5% for *Staphylococcus aureus* and *Escherichia coli*, respectively. Studies suggest that the use of rosemary oil and basil or rosemary oil and amyloglycosides was effective against *E. coli* (Barreto *et al.*, 2014; Sienkiewicz *et al.*, 2013). This oil showed potent control over Gram-positive bacteria than the Gram-negative bacteria. Higher minimum inhibition concentration (MIC), 2-5 μ g/ml of rosemary oil was observed in gram negative bacteria which indicates lesser colony control upon treatment (Moreno *et al.*, 2006; Hussain *et al.*, 2010; Jafari-sales and pashazadeh, 2020). Antibacterial activity with different polar extracts was tested and they proved to be effective for all the bacteria except *Bhrocotrix* sp. (Fernández-López *et al.*, 2005). Antibacterial activity was caused by organelles damage, increase in cell elasticity, vacuole segregations, increase in ROS in cells, depolarization of mitochondria, and cell membrane damage leading to lyses (Stojanović-Radić *et al.*, 2010; Shahina *et al.*, 2022).

Rosemary extracts showed antibacterial activities against *Shigella sonnei* and *Salmonella typhi* (Bozin *et al.*, 2007). Maximum

antibacterial activity was seen against *Rodotorula species* and *Scytalidium dimidiatum* through varied inhibition control mechanisms (Al-Bader, 2019) Rosemary oil extract at 100% concentration, showed inhibition of growth through clear areaformation in *Bacillus subtilis* (Elabass *et al.*, 2021). Inhibitory actions were more towards bacteria than yeasts at a minimal value of 0.5 to 1.5 mg/ml (Tavassoli *et al.*, 2011).

According to Gómez-Estaca *et al.* (2010), the essential oil of *R. officinalis* inhibited *Salmonella choleraesuis*, *S. sonnei*, *Yersinia enterocolitica*, *S. aureus*, *Bacillus cereus*, *Clostridium perfringens*, *Aeromonas hydrophila*, *Photobacterium phosphoreum* species growth completely. *Pseudomonas aeruginosa*, *Shewanella putrefaciens* growth were not inhibited.

5.2 Antifungal activity

The antifungal mechanism of rosemary extended towards fungal as well as yeast strains. This has proved to control *Aspergillus niger*, *Aspergillus fluves*, *Saccharomyces cervicise* and *Candidia albicans* (Abozid and Asker, 2013). Antagonistic activity on *C. albicans* was more predominant among the fungi group (Bozin *et al.*, 2007). Extract concentration from 60 to 80% had the maximum control over *C. albicans* (Matsuzaki *et al.*, 2013; Abozid and Asker, 2013). Individual treatment with cineole a major component showed control over the colonies (Matsuzaki *et al.*, 2013). The extract obtained in the later part of the distillation process, suppressed the growth in a better way (Genena *et al.*, 2008).

Twenty percent extract had an inhibition value of 32.31% against *A. niger*. The main antifungal activity was due to benzene and 1-methoxy-4-(1-propenyl) (Salih *et al.*, 2017). The suppression of the *Alternaria alternata* strain was due to the high 1,8-cineole content (Affes *et al.*, 2022). Mechanisms like physical changes in mycelium, increase in digestion activity of the cell wall, and changes in the

chemical composition of the cells, might be the reason for control in the growth of *Coletortichum gleosporoides* (Yuan *et al.*, 2024).

Extracts had shown great control over the plant pathogenic fungi also. Rosemary hydrosol was found to be an environmentally friendly antagonist source for *Fusarium* sp. as it inhibits growth up to 15% and does not show adverse effects on growth of plant species (Mravljje *et al.*, 2022). Essential oil combination of rosemary and sweet marjoram at a concentration of 100 and 125 and 15 µl/l of air hindered *Gleotrichum candidum* which causes potato rubbery rot disease (Kara, 2024).

6. Future prospects

In this growing world, importance is given majorly to efficiency and energy consumption of the process, due to growing demands all over the world. Hence, the right method of extraction for a better yield and quality oil will be an important criterion in the upcoming years. Exploration in the field of ethnopharmacology is on the run for the use of rosemary essential oil and the extracted constituents for the treatment of cancer as a main dose or supplementation is one of the interesting fields waiting for extensive research. Exploration for its use against various other pathogenic and non-pathogenic bacteria could be a great potent field to explore about. Hence, there are diverse fields to be explored, waiting for the scientific community to engage as an important medicinal and aromatic herb with its many hidden values.

7. Conclusion

Rosemary is an important crop medicinal plant division and has enormous potential in diverse fields. Though the usage of rosemary as a drug has been evolving, the basic requirements for deriving the compounds are the extraction procedures. Identification of the constituents with their respective class can give insight into the specification of these constituents. Emerging utilization of rosemary as a drug in pharmaceuticals for the supplementation in cancer and for the use as an antimicrobial agent is opening up new ventures. From this discussion, insights into the extraction methods, important constituents, and antimicrobial and anticancer properties have been provided as a framework.

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Conflict of interest

The authors declare no conflicts of interest relevant to this article.

Reference

Abu-Al-Basal, M. A. (2010). Healing potential of *Rosmarinus officinalis* L. on full-thickness excision cutaneous wounds in alloxan-induced-diabetic BALB/c mice. *J. Ethnopharmacology*, **131**(2):443-450.

Arafa, G. K. (2019). Process engineering extract the aromatic oil of the rosemary plant by steam distillation and hydrodistillation methods. *Misr J. Agrl. Eng.*, **36**(3):953-968.

Ai-Sereiti', M. R.; Abu-Amer, M. and Sen, P. (1999). Pharmacology of rosemary (*Rosmarinus officinalis* Linn.) and its therapeutic potentials. *Indian J. Exp. Biol.*, **37**(2):124-30.

Al-Bader, S. (2019). Antifungal activity of nine plant oils against local *Rhodotorula species* and *Scytalidium dimidiatum*. *Aalborg Academy J. Med. Sci.*, **2**(3):52-70.

Al-Hijazeen, M. and Al-Rawashdeh, M. (2019). Preservative effects of rosemary extract (*Rosmarinus officinalis* L.) on quality and storage stability of chicken meat patties. *Food Sci. Technol. (Brazil)*, **39**(1):27-34.

Ali, N. M. (2021). Phytochemical screening and *in vitro* antimicrobial and anticancer activities of different extracts of *Rosmarinus officinalis* (Rosemary): A comparative study. *Adv. Pharmacl. Pharm.*, **9**(3):45-55.

Al-Maharik, N.; Jaradat, N.; Hawash, M.; Al-Lahham, S.; Qadi, M.; Shoman, I.; Jaber, S.; Rahem, R. A.; Hussein, F. and Issa, L. (2022). Chemical composition, antioxidant, antimicrobial and anti-proliferative activities of essential oils of *Rosmarinus officinalis* from five different sites in Palestine. *Separations*, **9**(11):339

Almela, L.; Sánchez-Muñoz, B.; Fernández-López, J. A.; Roca, M. J. and Rabe, V. (2006). Liquid chromatographic-mass spectrometric analysis of phenolics and free radical scavenging activity of rosemary extract from different raw material. *J. Chromatogr. A*, **1120**(1-2):221-229

Alsamri, H.; Hasasna, H.; El, Baby, B.; Alneyadi, A.; Dhaheri, Y.; Al, Ayoub, M. A.; Eid, A. H.; Vijayan, R. and Itratni, R. (2021). Carnosol is a novel inhibitor of p300 acetyltransferase in breast cancer. *Front. Oncol.*, **11**:210.

Amin, T.; Naik, H. R. and Hussain, S. Z. (2017). Chemotyping the Essential Oil in Different Rosemary (*Rosmarinus officinalis* L.) Plants grown in Kashmir Valley. *Biosci. Biotechnol. Res. Asia*, **14**(3):1025-1031.

Humberto, M. B.; Edson, C. S. F.; Edeltrudes de O. L.; Henrique, D. M. C.; Maria, F. B.M; Cícera, C.A. T; Saulo, R. T; Juciane, V.R; Aislan, P.L. de Abreu; Maria do Carmo, G.L; Roger, W.G.O; Antonia M.G.L. and José A. D. L. (2014). Chemical composition and possible use as adjuvant of the antibiotic therapy of the essential oil of *Rosmarinus officinalis* L., *Ind. Crops Prod.*, **59**:290-294.

Basile, A.; Jiménez-Carmona, M. M. and Clifford, A. A. (1998). Extraction of Rosemary by superheated water. *J. Agric. Food Chem.*, **46**(12):5205-5209.

Beltrán-Villalobos, K. L.; Déciga-Campos, M.; Aguilar-Mariscal, H.; González-Trujano, M. E.; Martínez-Salazar, M. F.; Ramírez-Cisneros, M. de los Á.; Ríos, M. Y. and López-Muñoz, F. J. (2017). Synergistic antinociceptive interaction of *Syzygium aromaticum* or *Rosmarinus officinalis* coadministered with ketorolac in rats. *Biomed. Pharmacother.*, **94**:858-864.

Borges, R. S.; Ortíz, B. L. S.; Pereira, A. C. M.; Keita, H. and Carvalho, J. C. T. (2019). *Rosmarinus officinalis* essential oil: A review of its phytochemistry, anti-inflammatory activity, and mechanisms of action involved. *J. Ethnopharmacol.*, **229**:29-45.

Borrás-Linares, I.; Pérez-Sánchez, A.; Lozano-Sánchez, J.; Barrajón-Catalán, E.; Arráez-Román, D.; Cifuentes, A.; Micol, V. and Carretero, A. S. (2015). A bioguided identification of the active compounds that contribute to the antiproliferative/cytotoxic effects of rosemary extract on colon cancer cells. *Food Chem. Toxicol.*, **80**:215-222.

Bourhia, M.; Laasri, F. E.; Aourik, H.; Boukhris, A.; Ullah, R.; Bari, A.; Ali, S. S.; El Mzibri, M.; Benbacer, L. and Gmouh, S. (2019). Antioxidant and antiproliferative activities of bioactive compounds contained in *Rosmarinus officinalis* used in the mediterranean diet. *Evidence-Based Complementary and Alternative Medicine: eCAM*, **2019**: 7623830.

Bousbia, N.; Abert Vian, M.; Ferhat, M. A.; Petitcolas, E.; Meklati, B. Y. and Chemat, F. (2009). Comparison of two isolation methods for essential oil from rosemary leaves: Hydrodistillation and microwave hydrodiffusion and gravity. *Food Chem.*, **114**(1):355-362.

- Boutekdjret, C., Bentahar, F., Belabbes, R. and Bessiere, J. M. (2003). Extraction of rosemary essential oil by steam distillation and hydrodistillation. *Flavour Fragr J.*, **18**(6):481-484.
- Bozin, B.; Mimica-Dukic, N.; Samojlik, I. and Jovin, E. (2007). Antimicrobial and antioxidant properties of Rosemary and Sage (*Rosmarinus officinalis* L. and *Salvia officinalis*, Lamiaceae) essential oils. *J. Agric. Food Chem.*, **55**(19):7879-7885.
- Cattaneo, L.; Cicconi, R.; Mignogna, G.; Giorgi, A.; Mattei, M.; Graziani, G.; Ferracane, R.; Grosso, A.; Aducci, P.; Schininà, M. E. and Marra, M. (2015). Anti-proliferative effect of *Rosmarinus officinalis* L. extract on human melanoma A375 cells. *PLoS ONE*, **10**(7):e0132439
- Conde-Hernández, L. A.; Espinosa-Victoria, J. R.; Trejo, A. and Guerrero-Beltrán, J. (2017). CO₂-supercritical extraction, hydrodistillation and steam distillation of essential oil of rosemary (*Rosmarinus officinalis*). *J. Food Eng.*, **200**:81-86.
- de Macedo, L. M.; Dos Santos, É. M.; Militão, L.; Tundisi, L. L.; Ataíde, J. A.; Souto, E. B. and Mazzola, P. G. (2020). Rosemary (*Rosmarinus officinalis* L., syn. *Salvia rosmarinus* Spenn.) and its topical applications: A review. *Plants*, **9**(5):651 MDPI AG.
- Dilas, S., Knez, Ž.; Èetojević-Simin, D.; Tumbas, V.; Škerget, M.; Èanadanovića-Brunet, J. and Èetkovića, G. (2012). *In vitro* antioxidant and antiproliferative activity of three rosemary (*Rosmarinus officinalis* L.) extract formulations. *Int. J. Food Sci. Technol.*, **47**(10):2052-2062.
- Duran, T.; Koçak, N. and Karaselek, M. (2024). Carnosol and carnosic acid may be a promising anticancer agent in non-small cell lung cancer treatment. *Cukurova Med. J.*, **49**(1):81-88.
- El Boukhari, R. and Fatimi, A. (2023). Extract of rosemary as food additive: The landmark patents. *Biol. Life Sci. Forum*, **26**(1):37.
- Elabass, R. E. A.; Abdu, H.; Osman, H.; Ali, E.; Elkhilil, I.; Ali, R. E. A.; Osman, H. A. H. and Elkhilil, E. A. I. (2021). Impact of Rosemary (*Rosmarinus officinalis*) oil extract on microbial growth of three types of bacteria and yeast. *Asian Res. J. Current Sci.*, **3**(1):64-69.
- Elbanna, K.; Assiri, A. M. A.; Tadros, M.; Khider, M.; Assaedi, A.; Mohdaly, A. A. A. and Ramadan, M. F. (2018). Rosemary (*Rosmarinus officinalis*) oil: Composition and functionality of the cold-pressed extract. *J. Food Meas. Charact.*, **12**(3):1601-1609.
- Elyemni, M.; Louaste, B.; Nechad, I.; Elkamli, T.; Bouia, A.; Taleb, M.; Chaouch, M. and Eloutassi, N. (2019). Extraction of essential oils of *Rosmarinus officinalis* L. by two different methods: Hydrodistillation and microwave assisted Hydrodistillation. *Sci. World J.*, 2019.
- Fazlali, A.; Moradi, S. and Hamed, H. (2015). Studying of optimization condition of rosemary essence extraction with microwave assisted hydro-distillation method. *Am. J. Essent. Oil. Nat. Prod.*, **46**(1):46-50.
- Fernández-López, J.; Zhi, N.; Aleson-Carbonell, L.; Pérez-Alvarez, J. A. and Kuri, V. (2005). Antioxidant and antibacterial activities of natural extracts: Application in beef meatballs. *Meat Sci.*, **69**(3):371-380.
- Ferreira, D. F.; Lucas, B. N.; Voss, M.; Santos, D.; Mello, P. A.; Wagner, R.; Cravotto, G. and Barin, J. S. (2020). Solvent-free simultaneous extraction of volatile and non-volatile antioxidants from rosemary (*Rosmarinus officinalis* L.) by microwave hydrodiffusion and gravity. *Ind. Crop. Prod.*, **145**:112094
- Filly, A.; Fernandez, X.; Minuti, M.; Visinoni, F.; Cravotto, G. and Chemat, F. (2014). Solvent-free microwave extraction of essential oil from aromatic herbs: From laboratory to pilot and industrial scale. *Food Chem.*, **150**:193-198.
- Gachkar, L.; Yadegari, D.; Rezaei, M. B.; Taghizadeh, M.; Aastaneh, S. A. and Rasooli, I. (2007). Chemical and biological characteristics of *Cuminum cyminum* and *Rosmarinus officinalis* essential oils. *Food Chem.*, **102**(3):898-904.
- Gómez-Estaca, J.; López de Lacey, A.; López-Caballero, M. E.; Gómez-Guillén, M. C. and Montero, P. (2010). Biodegradable gelatin-chitosan films incorporated with essential oils as antimicrobial agents for fish preservation. *Food Microbiol.*, **27**(7):889-896.
- González-Vallinas, M.; Molina, S.; Vicente, G.; Zarza, V.; Martín-Hernández, R.; García-Risco, M. R.; Fornari, T.; Reglero, G. and De Molina, A. R. (2014). Expression of MicroRNA-15b and the glycosyltransferase GCNT3 correlates with antitumor efficacy of rosemary diterpenes in colon and pancreatic cancer. *PLoS ONE*, **9**(6):e98556
- GratiAffes, T.; Lasram, S.; Hammami, M.; Yeddes, W.; Wannes, W. A.; Khammassi, S.; Nasraoui, B.; Tounsi, M. S. and Hmida, N. L. Ben. (2022). A comparative assessment of antifungal activity of essential oils of five medicinal plants from Tunisia. *Int. J. Plant Based Pharm.*, **2**(2):220-227.
- Guo, Y.; Xie, J.; Li, X.; Yuan, Y.; Zhang, L.; Hu, W.; Luo, H.; Yu, H. and Zhang, R. (2018). Antidepressant effects of rosemary extracts associate with anti-inflammatory effect and rebalance of gut microbiota. *Front. Pharmacol.*, **9**:eCollection 2018.
- Habil Akos Mathe. (2018). Remarks to the recent trends in exploring medicinal plant diversity. *Ann. Phytomed.*, **7**(2):1-5(2018)
- Habtariam, S. (2023). Anti-inflammatory therapeutic mechanisms of natural products: Insight from rosemary diterpenes, carnosic acid and carnosol. *Biomedicines*, **11**(2):545
- Hcini, K.; Sotomayor, J. A.; Jordan, M. J. and Bouzid, S. (2013). Chemical composition of the essential oil of rosemary (*Rosmarinus officinalis* L.) of Tunisian origin. *Asian J. Chem.*, **25**(5):2601-2603.
- Hosni, K.; Hassen, I.; Chaâbane, H.; Jemli, M.; Dallali, S.; Sebei, H., and Casabianca, H. (2013). Enzyme-assisted extraction of essential oils from thyme (*Thymus capitatus* L.) and rosemary (*Rosmarinus officinalis* L.): Impact on yield, chemical composition and antimicrobial activity. *Ind. Crop. Prod.*, **47**:291-299.
- Huang, Y.; Xu, H.; Ding, M.; Li, J.; Wang, D.; Li, H.; Sun, M.; Xia, F.; Bai, H.; Wang, M.; Mo, M. and Shi, L. (2023). Screening of rosemary essential oils with different phytochemicals for antioxidant capacity, keratinocyte cytotoxicity, and anti-proliferative activity. *Molecules*, **28**(2):586.
- Hussain, A. I.; Anwar, F.; Chatha, S. A.; Jabbar, A.; Mahboob, S. and Nigam, P. S. (2010). *Rosmarinus officinalis* essential oil: antiproliferative, antioxidant and antibacterial activities. *Braz. J. Microbiol.*, **41**(4):1070-1078
- Jafari-sales, A. and Pashazadeh, M. (2020). Study of chemical composition and antimicrobial properties of Rosemary (*Rosmarinus officinalis*) essential oil on *Staphylococcus aureus* and *Escherichia coli* in vitro. *Int. J. Life Sci. Biotechnol.*, **3**(1):62-69.
- Jaglanian, A.; Termini, D. and Tsiani, E. (2020). Rosemary (*Rosmarinus officinalis* L.) extract inhibits prostate cancer cell proliferation and survival by targeting Akt and mTOR. *Biomed. Pharmacother.*, **131**:110717.
- Jaglanian, A. and Tsiani, E. (2020). Rosemary extract inhibits proliferation, survival, akt, and mTOR signaling in triple-negative breast cancer cells. *Int. J. Mol. Sci.*, **21**(3):810.
- Jang, Y. G.; Hwang, K. A. and Choi, K. C. (2018). Rosmarinic acid, a component of rosemary tea, induced the cell cycle arrest and apoptosis through modulation of HDAC2 expression in prostate cancer cell lines. *Nutrients.*, **10**(11):1784.
- Johnson, J. J.; Syed, D. N.; Heren, C. R.; Suh, Y.; Adhami, V. M. and Mukhtar, H. (2008). Carnosol, a dietary diterpene, displays growth inhibitory effects in human prostate cancer PC3 cells leading to G2-phase cell cycle arrest and targets the 52 -AMP-activated protein kinase (AMPK) pathway. *Pharm. Res.*, **25**(9):2125-2134.

- Kabubii, Z. N.; Mbaria, J. M.; Mathiu, P. M.; Wanjohi, J. M. and Nyaboga, E. N. (2024). Diet supplementation with rosemary (*Rosmarinus officinalis* L.) leaf powder exhibits an antidiabetic property in streptozotocin-induced diabetic male wistar rats. *Diabetology*, **5**(1):12-25.
- Adel, K.; Zied, Z.; Ines Ben, C.; Ahmed, B.; Neji, G.; Mohamed, D. and Gdoura, R. (2011). Chemical constituents and antioxidant properties of the essential oil from of *Rosmarinus officinalis* L. cultivated from the South-Western of Tunisia. *J. Med. Plant Res.*, **5**(25):5999-6004
- Kamal Genena, A.; Hense, H.; Smânia junior, A. and Machado de Souza, S. (2008). Rosemary (*Rosmarinus officinalis*): A study of the composition, antioxidant and antimicrobial activities of extracts obtained with supercritical carbon dioxide. *Ciênc. Tecnol. Aliment.*, **28**(2):463-469.
- Kara, M. (2024). Determination of chemical compositions of rosemary and sweet marjoram essential oils and their blends and their antifungal potential against potato rubbery rot disease agent *Geotrichum candidum*. *J. Plant Pathol.*, **106**:1173-1186
- Karakaya, S.; El, S. N.; Karagozlu, N.; Sahin, S.; Sumnu, G. and Bayramoglu, B. (2014). Microwave-assisted hydrodistillation of essential oil from rosemary. *J. Food Sci. Technol.*, **51**(6):1056-1065.
- Khademi Doozakhdarreh, S. F.; Khorshidi, J. and Morshedloo, M. R. (2022). Essential oil content and components, antioxidant activity and total phenol content of rosemary (*Rosmarinus officinalis* L.) as affected by harvesting time and drying method. *Bull. Natl. Res. Cent.*, **46**(1):511-527.
- Lorenzo, J. M.; Munekata, P. E. S.; Pateiro, M.; Dominguez, R.; Abdulrazzaq Alaghbari, M. and Tomasevic, I. (2021). Preservation of meat products with natural antioxidants from rosemary. *IOP Conf. Ser.: Earth Environ. Sci.*, **854**(1):317-332.
- Malekydozadeh, M.; Khadiv-Parsi, P.; Rezazadeh, S.; Abolghasemi, H.; Salehi, Z. and Li, Q. (2012). Application of multistage steam distillation column for extraction of essential oil of *Rosmarinus officinalis* L. *Iran. J. Chem. Eng.*, **9**(4):54-64.
- Manville, R. W.; Hogenkamp, D. and Abbott, G. W. (2023). Ancient medicinal plant rosemary contains a highly efficacious and isoform-selective KCNQ potassium channel opener. *Commun. Biol.*, **6**(1):644.
- Matsuzaki, Y.; Tsujisawa, T.; Nishihara, T.; Nakamura, M. and Kakinoki, Y. (2013). Antifungal activity of chemotype essential oils from rosemary against *Candida albicans*. *Open Access J. Stomatology*, **3**(2):176-182.
- Medhat Alzaharna, M.; El Hindi, M. W.; Sharif, F. A.; El-burai, H.; Alzaharna, M.; El-Hindi, M.; Sharif, F. and Isleem, R. (2020). Anticancer activity of Rosemary (*Rosmarinus officinalis* L.) and Oats (*Avena sativa* L.) Extracts, and their antitumor enhancement of 5-Fluorouracil on Colon Cancer Caco-2 Cell Line. *Int. J. Herb. Med.*, **8**(3):138-146.
- Mena, P.; Cirlini, M.; Tassotti, M.; Herrlinger, K. A.; Dall'Asta, C. and Del Rio, D. (2016). Phytochemical profiling of flavonoids, phenolic acids, terpenoids, and volatile fraction of a rosemary (*Rosmarinus officinalis* L.) extract. *Molecules*, **21**(11):1576.
- Mohamed Abdoul-latif, F.; Am, A. and Ainane, A. (2021). Chemical composition of bay laurel and rosemary essential oils from Morocco and their antifungal activity against fusarium strains. *Pharm. Online*, **2**:426-432.
- Moreno, S.; Scheyer, T.; Romano, C. S. and Vojnov, A. A. (2006). Antioxidant and antimicrobial activities of rosemary extracts linked to their polyphenol composition. *Free Radical Res.*, **40**(2):223-231.
- Mostafa Abozid, M. and Asker, M. (2013). Chemical composition, antioxidant and antimicrobial activity of the essential oil of the thyme and rosemary. *Int. J. Acad. Res.*, **5**(3):186-195.
- Mravlje, J.; Kopač, E.; Kosovel, H.; Leskošek, J. and Regvar, M. (2022). Potential of rosemary hydrosol for effective growth inhibition of fungi isolated from buckwheat grains. *Acta Biol. Slov.*, **65**(1):70-79.
- Mrđanović, J.; Bogdanović, V.; Kiprović, B.; Malenčič, D.; Mikulić-Petkovšek, M.; Milovanović, I. and Mišan, A. (2019) Muhammed Majeed. Novel insights to the anti-proliferative activity of rosemary (*Rosmarinus officinalis* L.) co-treatment. *Lekovite Sirovine*, **39**:44-51.
- Muhammed Majeed (2017). Evidence-based medicinal plant products for health care of world population. *Ann. Phytomed*, **6**(1):1-4.
- Mwithiga, G.; Maina, S.; Gitari, J. and Muturi, P. (2022). Rosemary (*Rosmarinus officinalis* L.) growth rate, oil yield and oil quality under differing soil amendments. *Heliyon*, **8**(4):920-921.
- Naimi, M.; Vlavcheski, F.; Shamshoum, H. and Tsiani, E. (2017). Rosemary extract as a potential anti-hyperglycemic agent: Current evidence and future perspectives. *Nutrients*, **9**(9):1-19.
- Nematollahi, P.; Mehrabani, M.; Karami-Mohajeri, S. and Dabaghzadeh, F. (2018). Effects of *Rosmarinus officinalis* L. on memory performance, anxiety, depression, and sleep quality in university students: A randomized clinical trial. *Complement. Ther. Clin. Pract.*, **30**:24-28.
- O'Neill, E. J.; Moore, J.; Song, J. and Tsiani, E. L. (2022). Inhibition of non-small cell lung cancer proliferation and survival by rosemary extract is associated with activation of ERK and AMPK. *Life*, **12**(1):52.
- O'Neill, E. J.; Sze, N. S. K.; MacPherson, R. E. K. and Tsiani, E. (2024). Carnosic acid against lung cancer: Induction of autophagy and activation of sestrin-2/LKB1/AMPK signalling. *Int. J. Mol. Sci.*, **25**(4):1950.
- Pawłowska, K.; Janda, K. and Jakubczyk, K. (2020). Properties and use of rosemary (*Rosmarinus officinalis* L.). *Pomeranian J. Life Sci.*, **66**(3):76-82.
- Pérez-Sánchez, A.; Barrajón-Catalán, E.; Ruiz-Torres, V.; Agulló-Chazarra, L.; Herranz-López, M.; Valdés, A.; Cifuentes, A. and Micol, V. (2019). Rosemary (*Rosmarinus officinalis*) extract causes ROS-induced necrotic cell death and inhibits tumor growth in vivo. *Sci. Rep.*, **9**(1):808.
- Pintore, G.; Usai, M.; Bradesi, P.; Juliano, C.; Boatto, G.; Tomi, F.; Chessa, M.; Cerri, R. and Casanova, J. (2002). Chemical composition and antimicrobial activity of *Rosmarinus officinalis* L. oils from Sardinia and Corsica. *Flavour Fragr. J.*, **17**(1):15-19.
- Presti, M. L., Ragusa, S.; Trozzi, A.; Dugo, P.; Visinoni, F.; Fazio, A.; Dugo, G. and Mondello, L. (2005). A comparison between different techniques for the isolation of rosemary essential oil. *J. Sep. Sci.*, **28**(3):273-280.
- Rahbardar, M. G. and Hosseinzadeh, H. (2020). Therapeutic effects of rosemary (*Rosmarinus officinalis* L.) and its active constituents on nervous system disorders. *Iran. J. Basic Med. Sci.*, **23**(9):1100-1112.
- Rajeswara Rao, B.; Kumar Kothari, S. and Pandu Sastry, K. (2010). Composition of rosemary (*Rosmarinus officinalis* L.) oil produced from the semi-arid tropical climate of south India. *J. Med. Aromat. Plant Sci.*, **32**:20-23.
- Reema, R.; Shashi, J. and Mamta, K. (2017). Scientific validation of traditional wisdom on analgesic effect of selected plant source. *Ann. Phytomed*, **6**(1):1-13.
- Rezzoug, S. A.; Baghdadi, M. W.; Louka, N.; Boutekedjiret, C. and Allaf, K. (1998). Study of a new extraction process: Controlled instantaneous decompression. Application to the extraction of essential oil from rosemary leaves. *Flavour Frag. J.*, **13**(4):251-258.
- Rezzoug, S. A.; Boutekedjiret, C. and Allaf, K. (2005). Optimization of operating conditions of rosemary essential oil extraction by a fast controlled pressure drop process using response surface methodology. *J. Food Eng.*, **71**(1):9-17.

- Sadeh, D.; Nitzan, N.; Chaimovitch, D.; Shachter, A.; Ghanim, M. and Dudai, N. (2019). Interactive effects of genotype, seasonality and extraction method on chemical compositions and yield of essential oil from rosemary (*Rosmarinus officinalis* L.). *Ind. Crops Prod.*, **138**(5):1114-19.
- Saini, A.; Pandey, A.; Sharma, S.; Suradkar, U. S.; Ambedkar, Y. R.; Meena, P.; Raman, R. and Gurjar, A. S. (2020). Assessment of antioxidant activity of rosemary (*Rosmarinus officinalis*) leaves extract. *Journal of Pharmacogn. Phytochem.*, **9**(3):14-17.
- Salih, T. F. M.; Mohammed, L. M. and Qader, K. O. (2017). Chemical analysis and growth inhibitory effect of rosemary plant on *Aspergillus niger*. *Kurdistan J. Appl. Res.*, **2**(2):151-154.
- Shahina, Z.; Homsy, R. Al; Price, J. D. W.; Whiteway, M.; Sultana, T. and Dahms, T. E. S. (2022). Rosemary essential oil and its components 1,8-cineole and α -pinene induce ROS-dependent lethality and ROS-independent virulence inhibition in *Candida albicans*. *PLoS ONE*, **17**(11): eCollection 2022.
- Sienkiewicz, M.; Łysakowska, M.; Pastuszka, M.; Bienias, W. and Kowalczyk, E. (2013). The potential of use basil and rosemary essential oils as effective antibacterial agents. *Molecules*, **18**(8):9334-9351.
- Socaci, S. A.; Tofană, M.; Socaciu, C. and Semeniciu, C. (2009). Optimization of HS/GC-MS method for the determination of volatile compounds from indigenous rosemary. *J. Agroalim. Technol.*, **15**(1):45-49.
- Sokhibjon, A.; Dilfuza, J.; Jitendr, M.; Krishnendra Singh, N.; Megha, B. and Baljeet, S. S. (2024). Exploring fungal endophytes in a medicinal plant, *Lavandula officinalis* L.: Isolation, characterization, and plant growth-promoting potential. *Ann. Phytomed*, **13**(2):979-986.
- Spadi, A.; Angeloni, G.; Guerrini, L.; Corti, F.; Michelozzi, M.; Cencetti, G.; Parenti, A. and Masella, P. (2021). Using a plackett–burman design to maximize yield of rosemary essential oil by distillation. *Ind. Crops Prod.*, **166**(6):113488.
- Stojanović-Radić, Z.; Nešić, M.; Ćomija, L. and Radulović, N. (2010). Antimicrobial activity and cytotoxicity of commercial rosemary oil (*Rosmarinus officinalis* L.). *Biol. Nyssana.*, **1**:83-88.
- Tai, J.; Cheung, S.; Wu, M. and Hasman, D. (2012). Antiproliferation effect of Rosemary (*Rosmarinus officinalis*) on human ovarian cancer cells *in vitro*. *Phytomedicine*, **19**(5):436-443.
- Tavassoli, S. K.; Mousavi, S. M.; Emam-Djomeh, Z. and Razavi, S. H. (2011). Chemical composition and evaluation of antimicrobial properties of *Rosmarinus officinalis* L. essential oil. *Afr. J. Biotechnol.*, **10**(63):13895-13899.
- Tigrine-Kordjani, N.; Meklati, B. Y.; Chemat, F. and Guezil, F. Z. (2012). Kinetic investigation of rosemary essential oil by two methods: Solvent-free microwave extraction and hydrodistillation. *Food Anal. Methods*, **5**(3):596-603.
- Tomi, K.; Kitao, M.; Konishi, N.; Murakami, H.; Matsumura, Y. and Hayashi, T. (2016). Enantioselective GC-MS analysis of volatile components from rosemary (*Rosmarinus officinalis* L.) essential oils and hydrosols. *Bioscience, Biotechnol. Biochem.*, **80**(5):840-847.
- Tong, X. P.; Ma, Y. X.; Quan, D. N.; Zhang, L. Yan, M. and Fan, X. R. (2017). Rosemary extracts upregulate Nrf2, Sestrin2, and MRP2 protein level in human hepatoma HepG2 cells. *Evidence-Based Complementary and Alternative Medicine: eCAM*, **2017**:7359806
- Tschiggerl, C. and Bucar, F. (2010). Investigation of the volatile fraction of rosemary infusion extracts. *Sci. Pharma.*, **78**(3):483-492.
- Volza Grow Global.(2024). Rosemary exports from India: Market Size & Demand based on Export Trade Data.
- Wang, W.; Li, N.; Luo, M.; Zu, Y. and Efferth, T. (2012). Antibacterial activity and anticancer activity of *Rosmarinus officinalis* L. essential oil compared to that of its main components. *Molecules*, **17**(3):2704-2713.
- Wang, Z. Y.; Ferraro, T.; Huang, M.T.; Ho, C.T. and Wang, Y. (1994). Inhibition of skin tumorigenesis by rosemary and its constituents carnosol and ursolic acid, *American Association for Cancer Research: AACR*, **54**(3):701-708.
- Yeddes, W.; Hammami, M.; Khammassi, S.; Grati, A.T ; Aidi, W.W. and Saidini - Tounsi Moufida. (2022). Antibacterial activities of rosemary (*Rosmarinus officinalis*) essential oil and ethanol extract. *Open Access Res. J. Multidiscip. Studies*, **3**(1):001-008.
- Yuan, T.; Hua, Y.; Zhang, D.; Yang, C.; Lai, Y.; Li, M.; Ding, S.; Li, S. and Chen, Y. (2024). Efficacy and antifungal mechanism of rosemary essential oil against *Colletotrichum gloeosporioides*. *Forests*, **15**(2):377.
- Zaouali, Y.; Chograni, H.; Trimech, R.; Boussaid, M. (2013). Changes in essential oil composition and phenolic fraction in *Rosmarinus officinalis* L. var. *typicus* Batt. organs during growth and incidence on the antioxidant activity, *Ind. Crops Prod.*, **43**:412-419.

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